



1-800-533-1710

Custom Gene Panel, Hereditary

CGPH

PATIENT NAME TESTRNV, IMPLEMENTATION				ORDER NUMBER N029000182
PATIENT ID X100420561	DATE OF BIRTH 02/19/1968	AGE 55 Y	SEX Male	REQUESTED BY CLIENT TESTING
COLLECTED 5/29/2023, 8:00 AM	RECEIVED 5/30/2023, 3:28 PM	REPORTED 6/19/2023, 11:09 AM		
The collected, received, and reported dates and times on the report are in the time zone of the performing location. 7028846 MCL RochesterCampus Rochester MN 55901				CLIENT ORDER NUMBER X100420561 CLIENT MRN 321

TEST DESCRIPTION

Custom gene testing for the ALDOB, GALE, GALK1, GALM and GALT genes

SPECIMEN

WB Whole Blood

RESULT SUMMARY

Pathogenic Variant Detected

RESULT

The following homozygous PATHOGENIC variant was detected:
GALT (NM_000155.4), chr9(GRCh37):g.34648167A>G, c.563A>G, p.Gln188Arg (p.Q188R)

INTERPRETATION

GALT c.563A>G (p.Q188R), PATHOGENIC
The homozygous c.563A>G (p.Q188R) missense variant in the GALT gene (MIM:606999) is an established pathogenic variant. Pathogenic variants in the GALT gene have been associated with autosomal recessive galactosemia (1). The detected variants are in trans (on opposite chromosomes). This result is supportive of a molecular diagnosis of galactosemia for this individual. Clinical correlation is recommended.

This result should be interpreted in the context of clinical findings, family history, and other laboratory testing.

Consultation with a genetics professional is recommended for interpretation of this result and to determine whether reproductive risk assessment and familial testing may be of benefit to this family. Genetic testing for family members is available by ordering FMTT / Familial Mutation, Targeted Testing for the specific variant(s) detected. Please contact the laboratory at 1-800-533-1710 or the online test catalog at www.mayocliniclabs.com for information about FMTT.

REFERENCES:

1) GeneReviews: Classic Galactosemia and Clinical Variant Galactosemia
(<https://www.ncbi.nlm.nih.gov/books/NBK1518/>) (PMID: 20301691)

METHOD

Next generation sequencing (NGS) and/or Sanger sequencing was performed to test for the presence of variants in coding regions and intron/exon boundaries of the genes analyzed, as well as some other regions that have known pathogenic variants. NGS and/or a polymerase chain reaction (PCR)-based quantitative method was performed to test for the presence of deletions and duplications in the genes analyzed. The human genome reference GRCh37/hg19 build



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was used for sequence read alignment. At least 99% of the bases are covered at a read depth over 30X. Sensitivity is estimated at above 99% for single nucleotide variants, above 94% for indels less than 40 base pairs (bp), above 95% for deletions up to 75 bp and insertions up to 47 bp. See the Genes Analyzed field for a list of genes tested.

There may be regions of genes that cannot be effectively evaluated for sequencing or deletion and duplication analysis as a result of technical limitations of the assay, including regions of homology, high GC content, and repetitive sequences. Confirmation of select reportable variants was performed by alternate methodologies based on internal laboratory criteria. See www.mayocliniclabs.com (TEST ID CGPH) for details regarding genes with regions not routinely covered.

GENES ANALYZED

ALDOB, GALE, GALK1, GALM and GALT

GENE LIST ID

IEMCP-6D1TTY

DISCLAIMER**Clinical Correlations**

An online research opportunity called GenomeConnect (genomeconnect.org), a project of ClinGen, is available for the recipient of this genetic test. This patient registry collects de-identified genetic and health information to advance the knowledge of genetic variants. Mayo Clinic is a collaborator of ClinGen. This may not be applicable for all tests.

If testing was performed because of a clinically significant family history it is often useful to first test an affected family member. Detection of a reportable variant(s) in an affected family member would allow for more informative testing of at risk individuals.

To discuss the availability of further testing options or for assistance in the interpretation of these results, Mayo Clinic Laboratory genetic counselors can be contacted at 1-800-533-1710.

Technical Limitations

Next generation sequencing may not detect all types of genomic variants. In rare cases, false negative or false positive results may occur. The depth of coverage may be variable for some target regions, but assay performance below the minimum acceptable criteria or for failed regions will be noted. Given these limitations, negative results do not rule out the diagnosis of a genetic disorder. If a specific clinical disorder is suspected, evaluation by alternative methods can be considered.

There may be regions of genes that cannot be effectively evaluated for sequencing or deletion and duplication analysis as a result of technical limitations of the assay, including regions of homology, high GC content, and repetitive sequences. Confirmation of select reportable variants was performed by alternate methodologies based on internal laboratory criteria.

Additionally, low level mosaic variants may not be detected.

This test is not designed to differentiate between somatic and germline variants. If there is a possibility that any detected variant is somatic, additional testing may be necessary to clarify the significance of results.



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Reclassification of Variants Policy

See www.mayocliniclabs.com (TEST ID CGPH) for information regarding the laboratory's policy for reclassification of variants.

Variant Evaluation

Variant curation is performed using published ACMG-AMP recommendations as a guideline. Other gene-specific guidelines may also be considered. Variants classified as benign or likely benign are not reported.

RELEASED BY

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